

BACTERIAL CELL WALL CONSTITUENTS. II.<sup>1</sup> SYNTHESIS OF O-  
(N-ACETYL- $\beta$ -MURAMYL-L-ALANYL-D-ISOGLUTAMINE)-(1 $\rightarrow$ 4)-N-ACETYL-D-GLUCOSAMINE

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SUMMARY

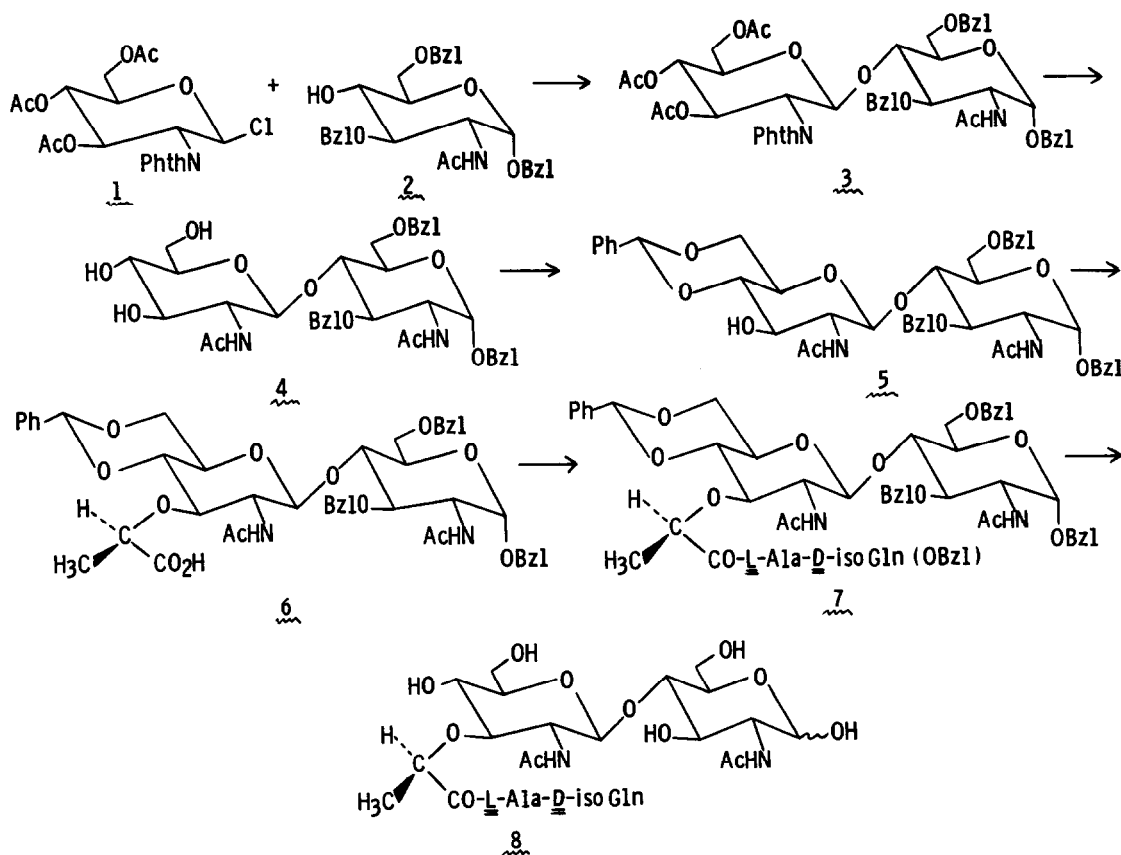
O-(N-Acetyl- $\beta$ -muramyl-L-alanyl-D-isoglutamine)-(1 $\rightarrow$ 4)-N-acetyl-D-glucosamine (8), the repeating disaccharide dipeptide unit obtained by endo-N-acetylglucosaminidase lysis of bacterial cell walls, has been synthesized in a twelve-step sequence from N-acetyl-D-glucosamine.

The glycodipeptide moiety of the rigid polymeric bacterial cell wall peptidoglycan is composed of alternating units of  $\beta$ 1 $\rightarrow$ 4 linked N-acetyl-D-glucosamine (GlcNAc) and N-acetyl-muramyl-L-alanyl-D-isoglutamine (muramyl dipeptide-MDP), MDP comprising the minimum structure<sup>2</sup> of mycobacteria in Freund's complete adjuvant necessary for immuneadjuvant activity. Therefore, two disaccharide dipeptide structures can be released by enzymic cleavage of the glycosidic linkages in the peptidoglycan, one [MDP-( $\beta$ 1 $\rightarrow$ 4)-GlcNAc] having N-acetyl-D-glucosamine and the other [GlcNAc-( $\beta$ 1 $\rightarrow$ 4)-MDP] muramyl dipeptide at the reducing end of the  $\beta$ 1 $\rightarrow$ 4 linked disaccharide. To assist in the elucidation of the minimal structural requirements for the arthritogenicity of peptidoglycans,<sup>3</sup> and, moreover, to provide compounds with potentially enhanced adjuvant properties over MDP, we have synthesized the two possible peptidoglycan isomers. In this communication we report the synthesis of MDP-( $\beta$ 1 $\rightarrow$ 4)-GlcNAc (8) in a twelve-step sequence from N-acetyl-D-glucosamine. Structure 8 represents the dipeptide derivative of the disaccharide [MurNAc-( $\beta$ 1 $\rightarrow$ 4)-GlcNAc] that is isolated from lysostaphin endo-N-acetylglucosaminidase digests of bacterial cell walls.<sup>4</sup> The synthesis of GlcNAc-( $\beta$ 1 $\rightarrow$ 4)-MDP will be described elsewhere.<sup>5</sup>

The strategy employed in the synthesis of 8 required the construction of an appropriately blocked  $\beta$ 1 $\rightarrow$ 4 linked disaccharide having only the C-3'<sup>6</sup> hydroxyl unprotected and thereby available for conversion to the (R)-lactic acid ether 6. This key intermediate (5) was obtained by condensation of the two D-glucosamine derivatives 1 and 2 and further chemical manipulation. The hydroxyls in the glucosaminyl donor 1 were blocked "temporarily"<sup>7</sup> as acetates, whereas those in the glucosaminyl acceptor 2 (free C-4 hydroxyl) "persistently"<sup>7</sup> as benzyl ethers and benzyl glycoside. The MurNAc-( $\beta$ 1 $\rightarrow$ 4)-GlcNAc derivative 6 was then prepared and coupled

with the requisite blocked dipeptide. Final deprotection of the resulting fully blocked disaccharide dipeptide 7 afforded desired 8. Protective groups had been chosen so that they could all be removed in a single hydrogenolytic step.

The synthesis of the blocked chitobiose derivative 3 was accomplished following the method developed by Lemieux and coworkers<sup>8</sup> for the preparation of 2-amino-2-deoxy- $\beta$ -glycopyranosides using 2-deoxy-2-phthalimidoglycosyl halides. Thus, reaction of 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido- $\beta$ -D-glucopyranosyl chloride (1)<sup>9</sup> with benzyl 2-acetamido-3,6-di-*O*-benzyl-2-deoxy- $\alpha$ -D-glucopyranoside (2)<sup>10</sup> in nitromethane in the presence of the silver triflate-collidine complex gave after column chromatography compound 3<sup>11</sup> in 40% yield;  $[\alpha]_{\text{D}}^{25} + 57^\circ$  ( $c$  1,  $\text{CHCl}_3$ ); n.m.r. (300-MHz,  $\text{CDCl}_3$ ):  $\delta$  1.74, 1.83, 1.94, and 1.99 (3-proton singlets, 3 OAc and 1 NHAc), 4.86 (d,  $\text{J}_{1,2}$  3.8 Hz, H-1), 5.15 (t,  $\text{J}_{3',4'}$  9.2 Hz, H-4'), 5.51 (d,  $\text{J}_{1',2'}$  8.5 Hz, H-1'), and 5.74 (dd,  $\text{J}_{2',3'}$  10.8 Hz, H-3'). De-*O*-acetylation and de-*N*-phthalylation of 3



Bzl = benzyl, Phth = phthaloyl, Ph = phenyl

was performed with *n*-butylamine in refluxing methanol (48h); subsequent *N*-acetylation with acetic anhydride in methanol afforded compound 4 (69% yield based on 3) having the C-3', C-4', and C-6' hydroxyls free; m.p. 252-256° (dec.) (ethanol),  $[\alpha]_{\text{D}}^{25} + 72^\circ$  (c 1.2, MeOH); n.m.r. (300-MHz, CD<sub>3</sub>OD):  $\delta$  1.94 and 1.97 (3-proton singlets, 2 NHAc), 4.82 (d,  $J_{1,2}$  3.9 Hz, H-1), and 5.06 (d,  $J_{1',2'}$  10.8 Hz, H-1'). The presence of a  $\beta(1\rightarrow4)$  linkage in 4 was confirmed by catalytic hydrogenolysis (Pd in AcOH) of the benzyl ether and benzyl glycoside protective groups to give di-*N*-acetylchitobiose identical in all respects (m.p.,  $[\alpha]_{\text{D}}$ , 300-MHz n.m.r., t.l.c. mobility) with an authentic sample obtained from chitin.<sup>12</sup> Blocking of the C-4' and C-6' hydroxyls in 4 as a benzylidene acetal (benzaldehyde-ZnCl<sub>2</sub>) provided the required intermediate 5 with only the C-3' hydroxyl unprotected; yield 90%, m.p. 298-302° (dec.) (methanol),  $[\alpha]_{\text{D}}^{25} + 58^\circ$  (c 1.1, DMF); n.m.r. (300-MHz, CDCl<sub>3</sub>):  $\delta$  1.77 and 1.82 (3-proton singlets, 2 NHAc), 4.93 (d,  $J_{1,2}$  3.8 Hz, H-1), and 5.47 (s, benzylic H). The (*R*)-lactic acid ether 6 was then obtained as an amorphous solid in 91% yield by alkylation<sup>13</sup> of 5 with (*S*)-2-chloropropionic acid in dioxane in the presence of sodium hydride;  $[\alpha]_{\text{D}}^{25} + 45^\circ$  (c 1, MeOH), n.m.r. (300-MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.12 [d, CH<sub>3</sub> (lac)], 1.82 (6-proton singlet, 2 NHAc), and 5.60 (s, benzylic H). The dipeptide was next introduced by the mixed anhydride method (reaction of acid 6 with *L*-alanyl-*D*-isoglutamine benzyl ester hydrochloride<sup>14</sup> in DMF in the presence of *N*-methylmorpholine and isobutyl chloroformate) to afford the fully blocked disaccharide dipeptide 7 in 93% yield; m.p. 295-300° (dec.),  $[\alpha]_{\text{D}}^{25} + 59^\circ$  (c 0.54, DMF); n.m.r. (300-MHz, DMSO-*d*<sub>6</sub>):  $\delta$  1.19 and 1.23 [2d, CH<sub>3</sub> (ala) and CH<sub>3</sub> (lac)], 1.80 and 1.84 (3-proton singlets, 2 NHAc), 2.34 (t, -CH<sub>2</sub>CO<sub>2</sub>Bzl), 5.07 (s, -CO<sub>2</sub>CH<sub>2</sub>Ph), and 5.66 (s, benzylic H). In the final step of the sequence, the benzyl ester, benzyl ether, benzyl glycoside, and benzylidene acetal protective groups were removed by hydrogenolysis of 7 in glacial acetic acid (Pd, 24 h, atmospheric pressure). Chromatographically homogeneous (silica gel t.l.c., 60:40:10 CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O) *O*-(*N*-acetyl- $\beta$ -muramyl-*L*-alanyl-*D*-isoglutamine)-(1 $\rightarrow$ 4)-*N*-acetyl-*D*-glucosamine (8) was isolated (precipitation by addition of diethyl ether to a methanol-ethanol solution) in 95% yield as an amorphous solid; n.m.r. (300-MHz, D<sub>2</sub>O):  $\delta$  1.38 and 1.45 [2d, CH<sub>3</sub> (ala) and CH<sub>3</sub> (lac)], 1.99 and 2.04 (3-proton singlets, 2 NHAc), 2.04 (m, -CH<sub>2</sub>CH<sub>2</sub>CO<sub>2</sub>H), and 2.29 (t, -CH<sub>2</sub>CO<sub>2</sub>H).

Compound 8 administered as an aqueous solution in mice significantly increased the level of antibodies against bovine serum albumin.<sup>15</sup> Details of its immunoadjuvant properties will be reported elsewhere.

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